



“The stress of shear”

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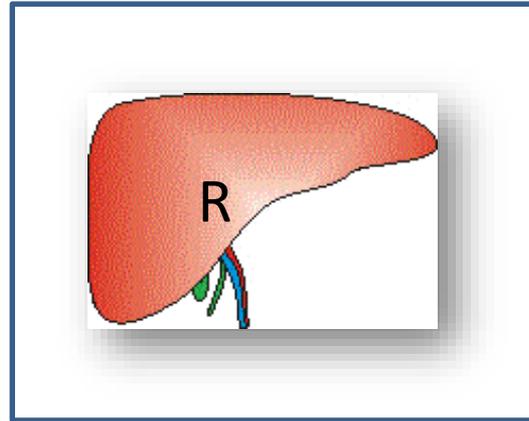
Centro E. Piaggio
bioengineering and robotics research center



Blood flow in, Q_{in}



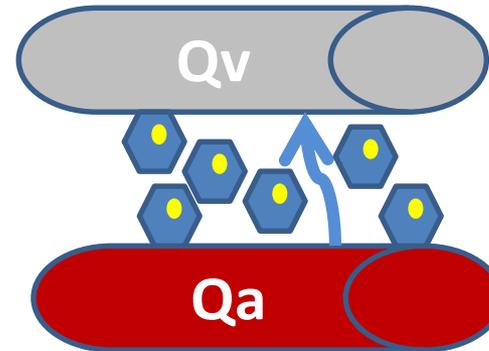
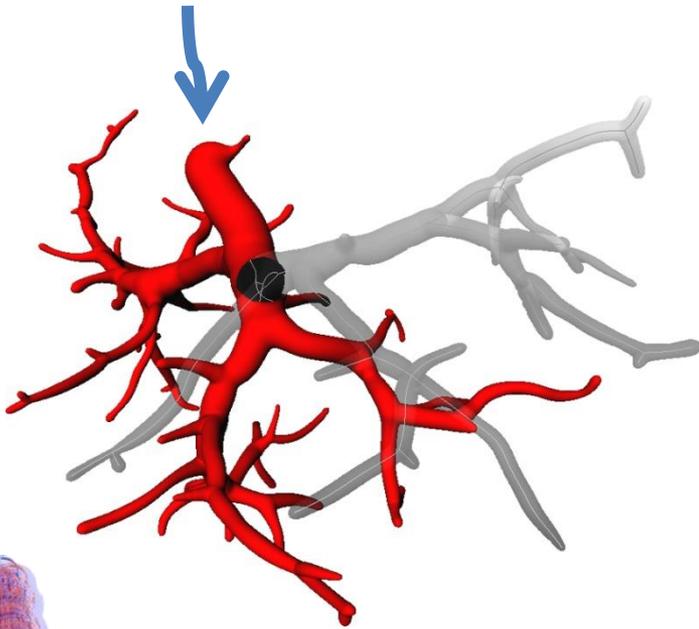
C_{in}



Blood flow out, Q_{out}



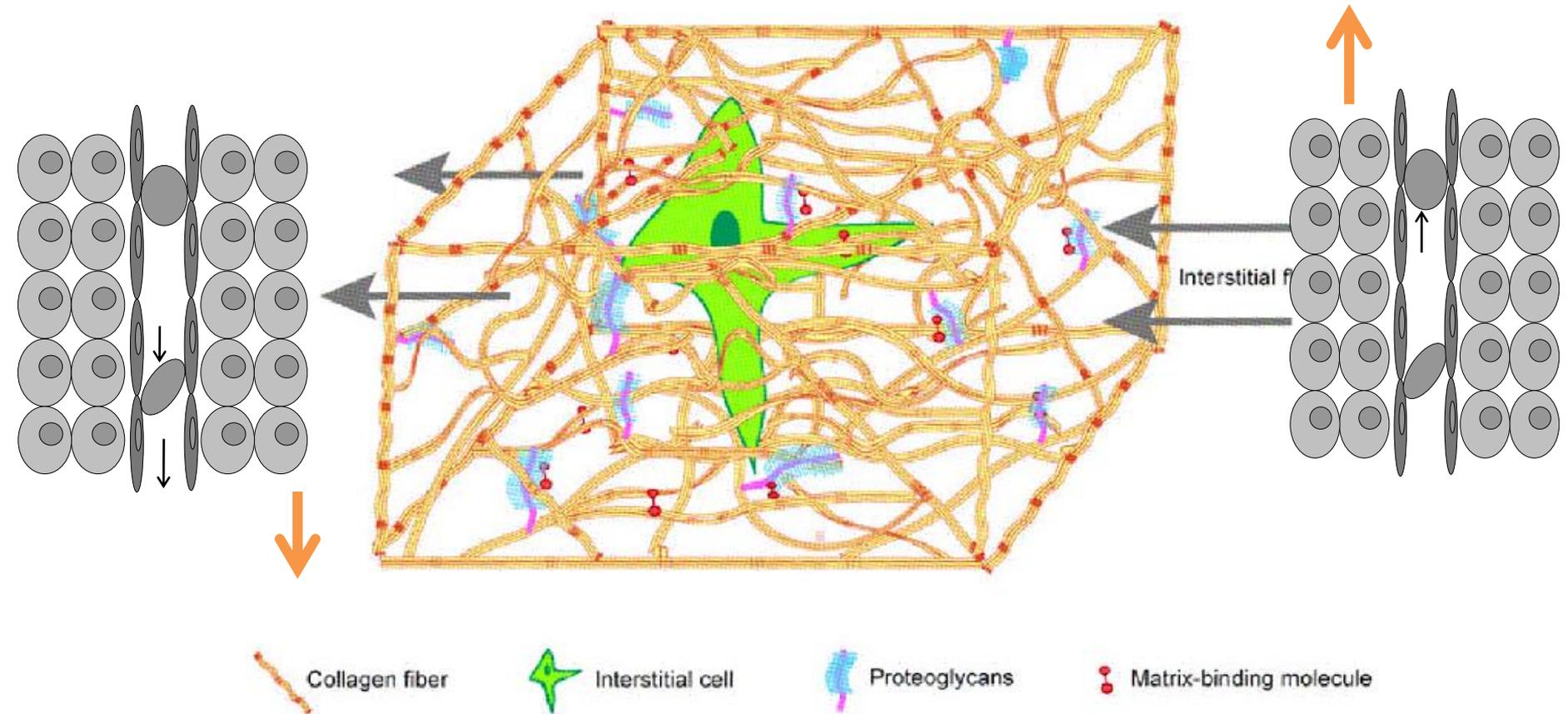
C_{out}



Interstitial
flow driven by
concentration
gradients



INTERSTITIAL FLOW



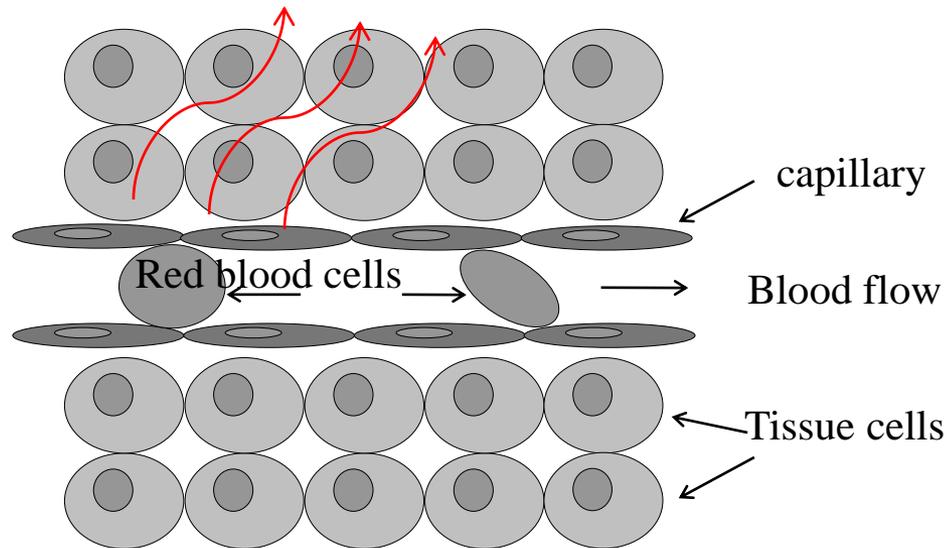
1) interstitial flow is due to a concentration gradient 2) all tissues are permeated by interstitial flow 3) the flow is through a microporous medium

Swartz & Fleury, ARBE
Vol. 9: 229-256.2007



FLOW and SHEAR

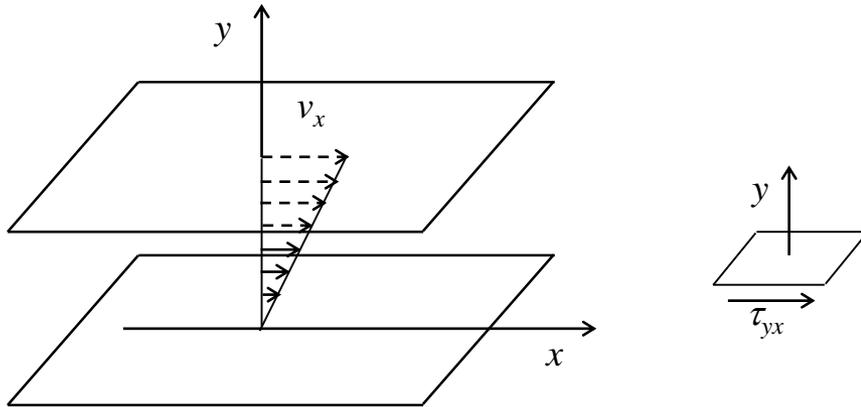
Only epithelial cells (skin, blood vessels, intestine) and the non adherent cells of the immune system and blood can support direct fluid flow.



The motion of fluid across a mobile or semi mobile surface gives rise to **shear stress**



Shear stress



$$\tau_{yx} = -\mu \frac{dv_x}{dy}$$

The shear stress on a monolayer of cells in a flat chamber with flow Q is

$$\tau_{yx} = -\frac{6Q\mu}{wh^2}$$

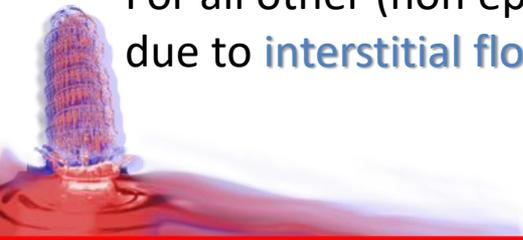


Optimal shear stress in bioreactors

Cell	Shear	Flow rate	Ref
Human trabecular bone, 3D	$5 \cdot 10^{-5}$ Pa	0.01 mL/min	Porter. Journal of Biomechanics, 38, 543, 2005
Human osteosarcoma cells, 3D	0-0.021 Pa	Max. 25 mL/min	Laganà. Biomedical Microdevices, 14(1), 225, 2012
hBMSC, 3D	0.015 Pa	3 mL/min	Li. Tissue Eng. A, 15, 2773, 2009
HepG2, 2D	0.14 Pa	0.0025 mL/min	Tanaka et al, Meas. Sci. Technol. 17, 3167–3170, 2006
Human hepatocytes, 2D+ gel	$5 \cdot 10^{-5}$ Pa	0.25 mL/min	Vinci et al. Biotech J., 6(5):554, 2011
Rat hepatocytes, 2D+ fibroblasts	0.014 Pa	0.06 mL/min	Tilles et al, Biotech & Bioeng. 73 (5), 379, 2001

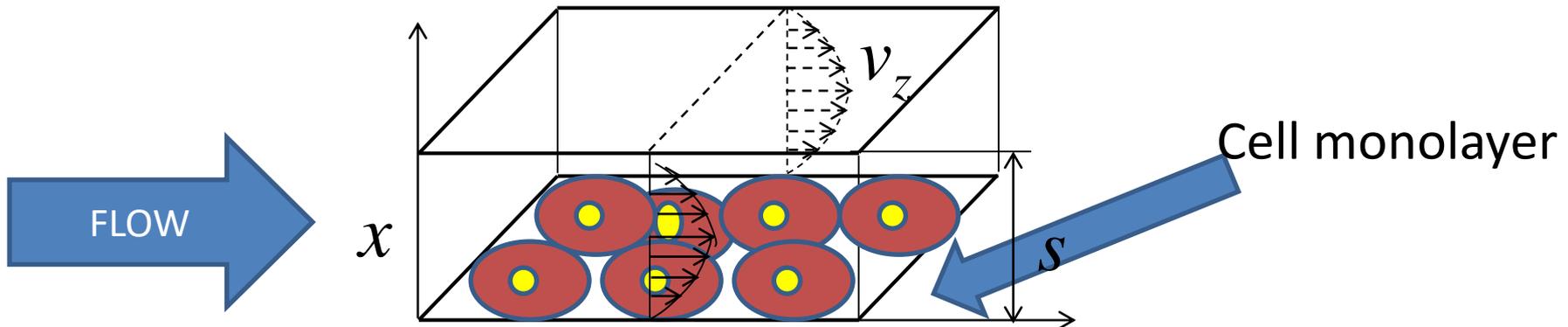
Wall shear stress in blood vessels: 1-0.01 N/m²

For all other (non epithelial) tissues shear is much less (0.01-0.00001 N/m²), and is due to **interstitial flow** (few microL/min).



Adding flow

$$\frac{dc}{dt} = D\nabla^2 c - R - v \cdot \nabla c$$



For a monolayer

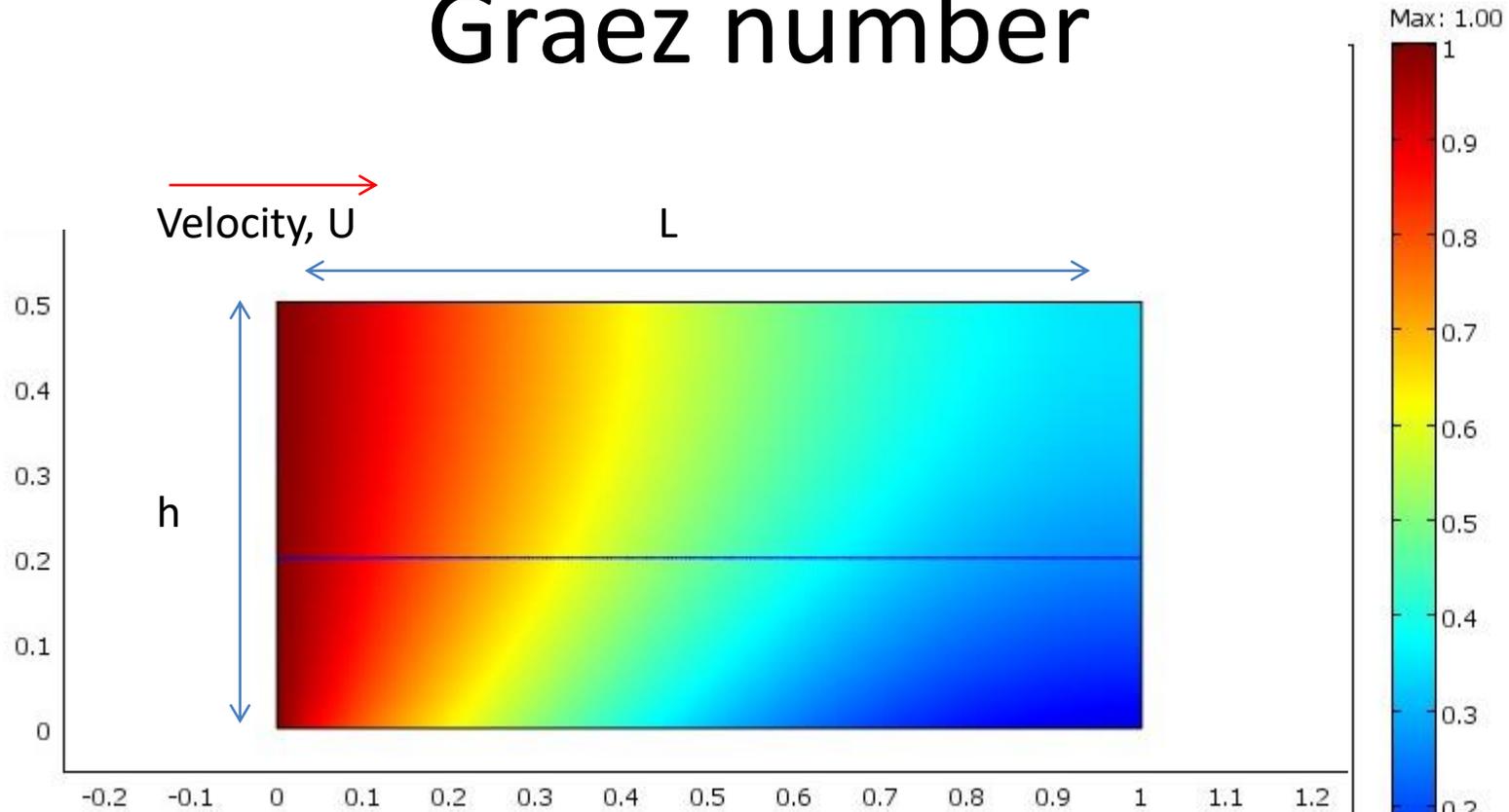
$$\frac{dc}{dt} = D \frac{dc^2}{dx^2} - v_z \frac{dc}{dz}$$

For volumetric consumption

$$\frac{dc}{dt} = D \frac{dc^2}{dx^2} - v_z \frac{dc}{dz} - R_{vol}$$



Graetz number

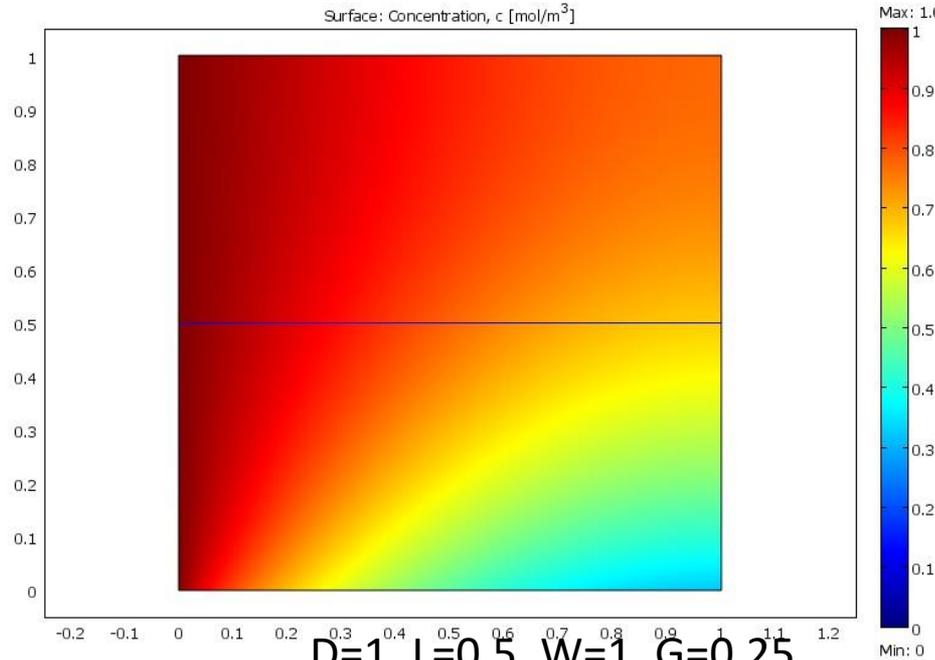


$G=0.25$

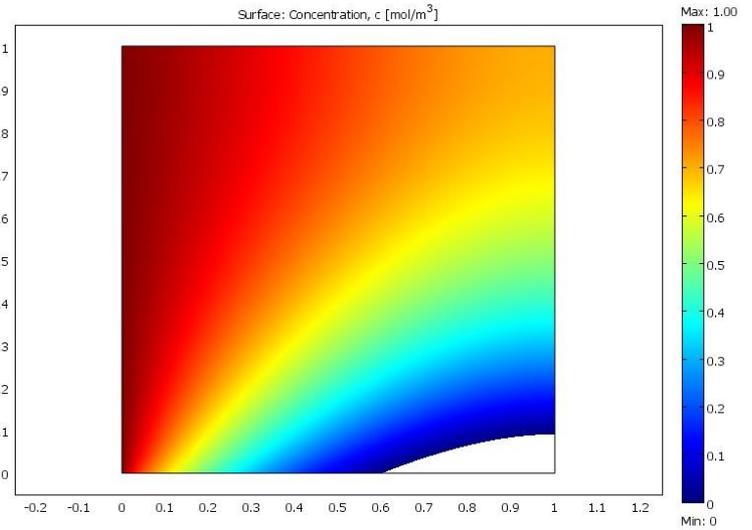
$$G = \frac{t_{diff}}{t_{conv}} = \frac{\frac{L^2}{D}}{\frac{h}{U}} = \frac{L^2 U}{Dh}$$

HOW CONCENTRATION PROFILES CHANGE WITH GRAEZ NUMBER

$D=1, L=1, W=1, G=1$

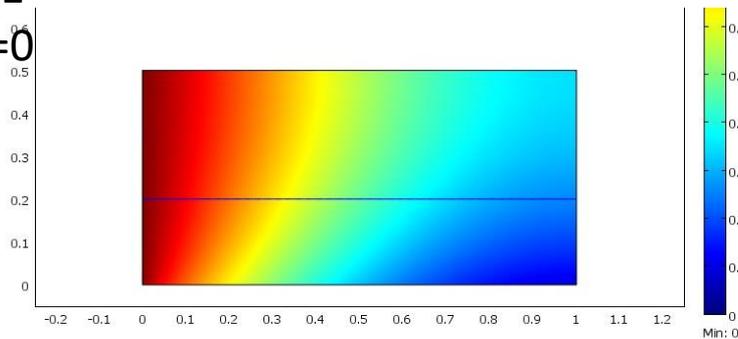


$D=2, L=1, W=1, G=0.5$



$D=1, L=0.5, W=1, G=0.25$

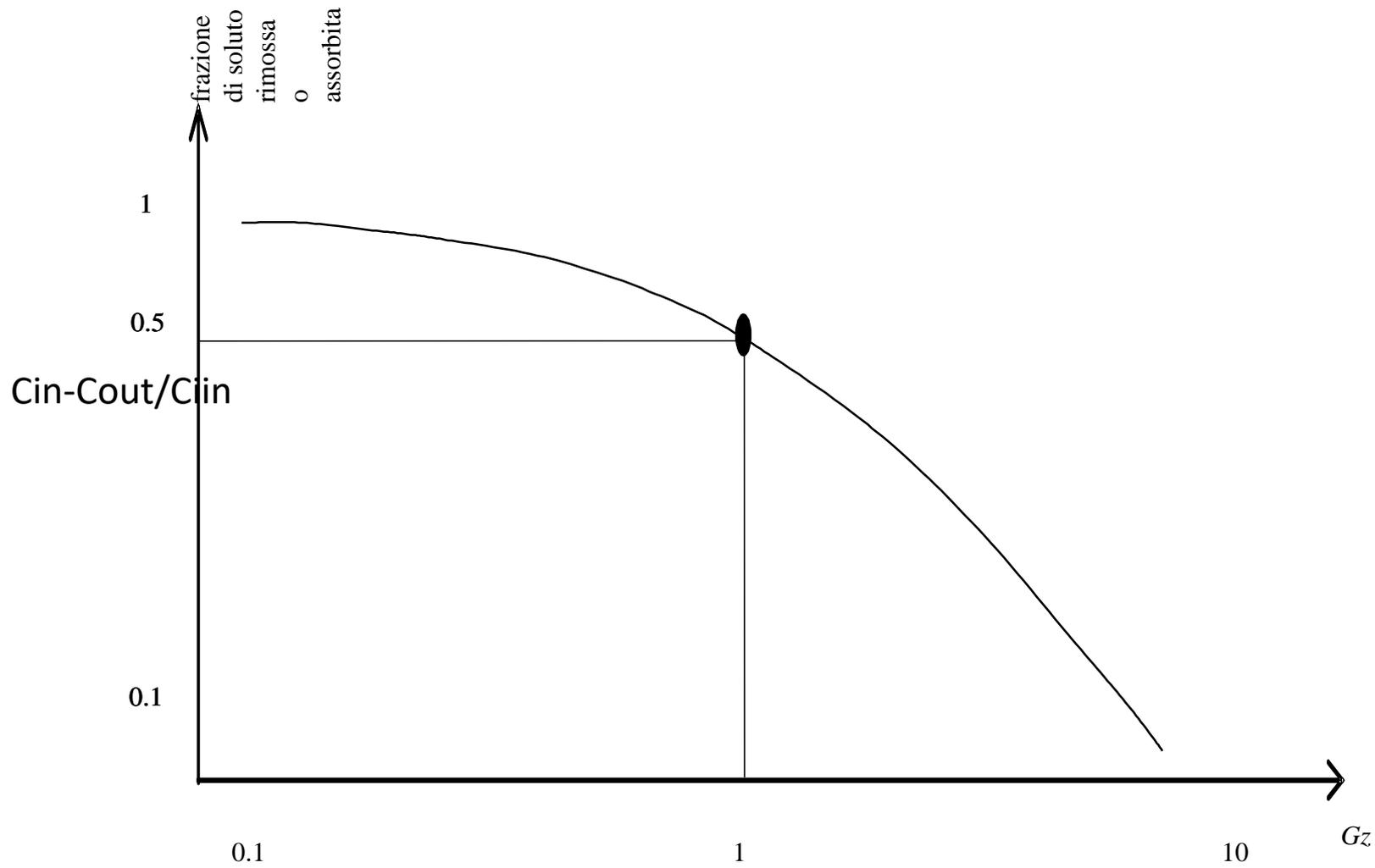
RED, $C=1$
BLUE $C=0$



$D=1, L=1, W=0.5, G=2$



0.4 0.5 0.6 0.7 0.8 0.9 1 1.1



Stop here

Example 1: Oxygen diffusion in gel encapsulated islets

100 μm
462
cells

150 μm
1560 cells

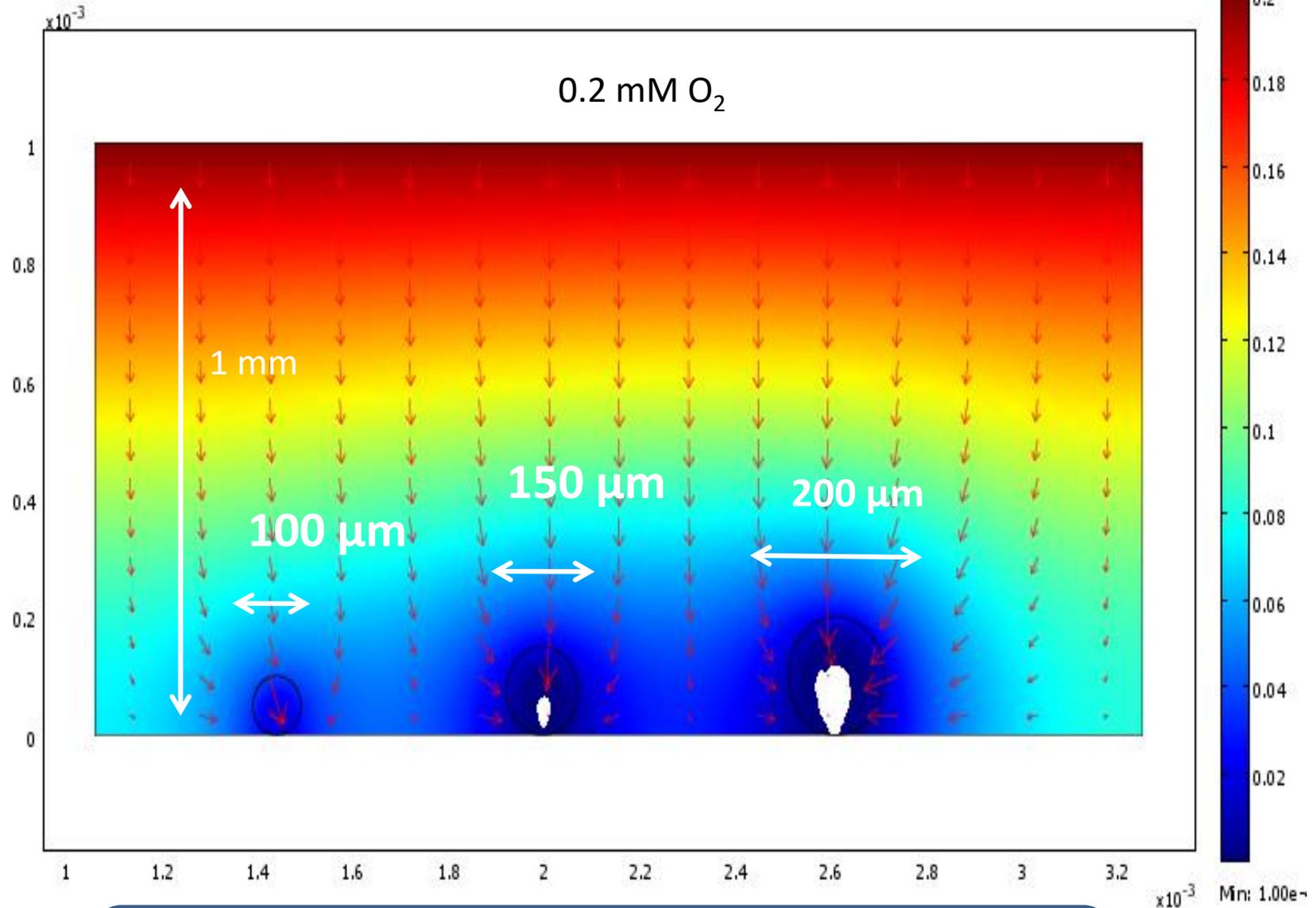
200 μm
3700 cells

Vmax	Km	C crit	Co	D in water	D in sphere
0.034 mM.s ⁻¹	1.10 ⁻³ mM (0.7 mmHg)	1.10 ⁻⁴ mM (0.07 mmHg)	0.2 mM	3.10 ⁻⁹ m ² .s ⁻¹	2.10 ⁻⁹ m ² .s ⁻¹
Medium height	δ (Heavyside)	Cell density			
1 mm	flc1hs(c- 0.02,0.0001)	8.8.10 ¹⁴ cells.m ⁻³			

$$R_{vol} = \frac{V_{max} c}{K_m + c} \cdot \delta$$



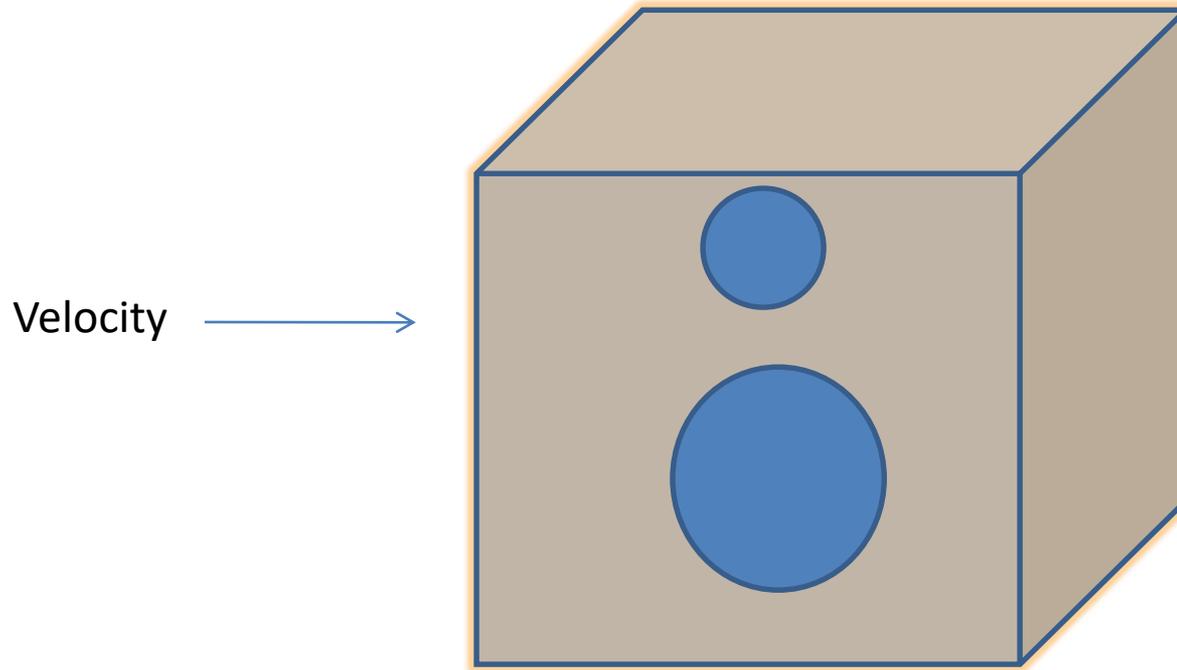
Surface: Concentration, c [mol/m³] Arrow: Diffusive flux, c



Simple problem solved with mass transfer equations in
Comsol Multiphysics



Example 2: Oxygen diffusion in perfused gel encapsulated islets in a bioreactor

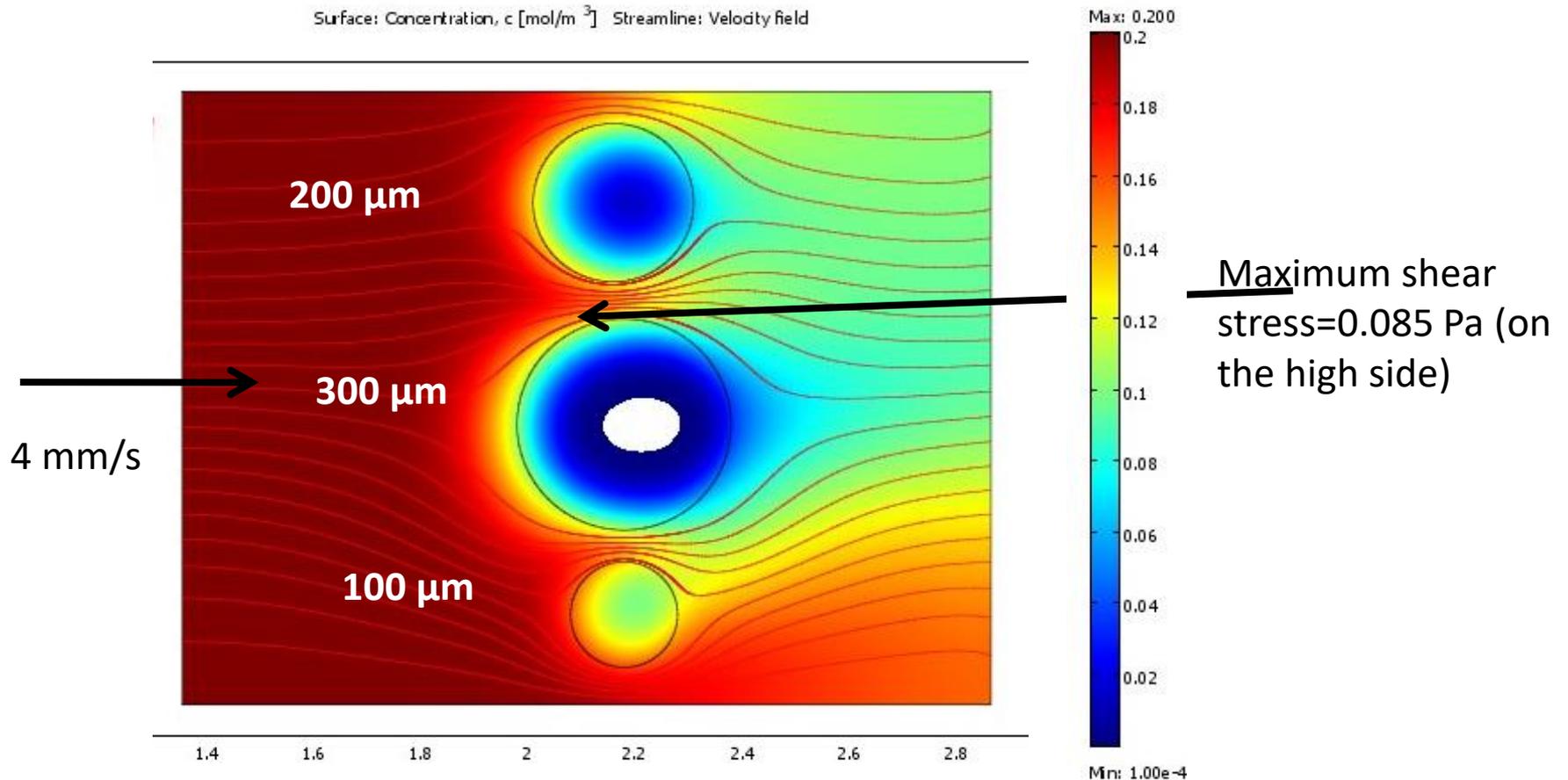


The islets are encapsulated in a non porous gel
Nutrients will only get to the cells by diffusion through the gel

Solved by coupling the Navier-Stokes equations for the fluidic domain with convection and diffusion



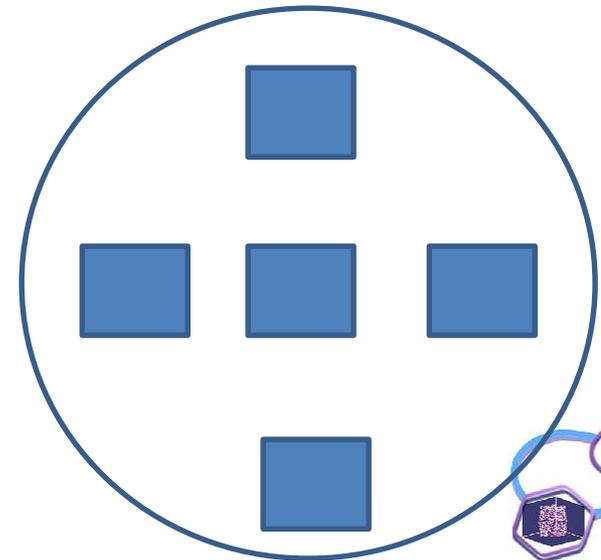
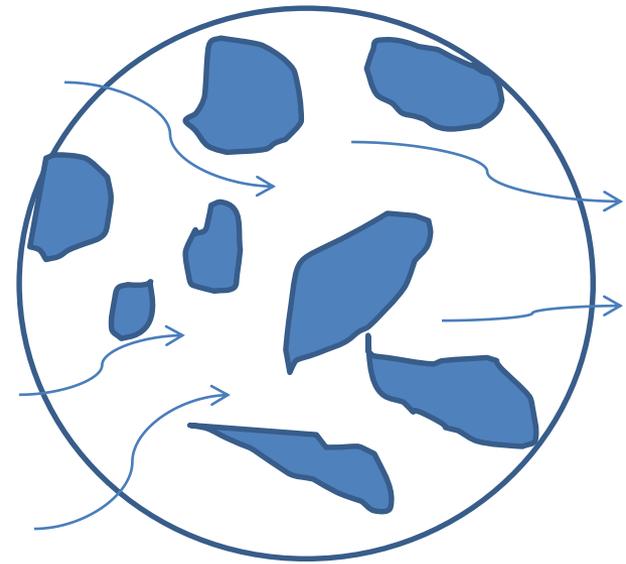
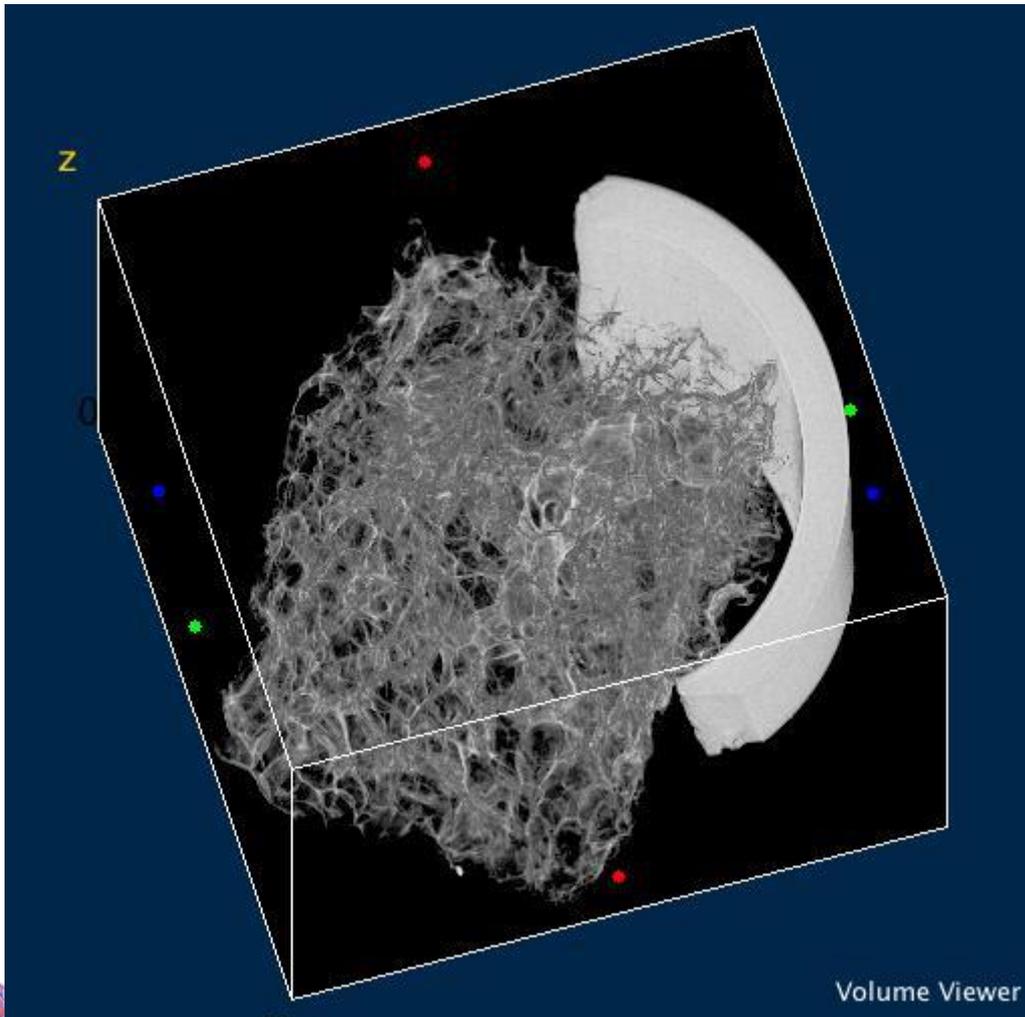
Islets in a bioreactor perfusion chamber, flow velocity of 4 mm/s.



The size limit is between 200 and 300 microns
- larger constructs have to be porous



Flow through pores



Darcy –Brinkman equations: enable calculation of average flow rate and shear in porous media, correlating pore level flow effects to the bulk fluid motion. In Darcy’s model, the average fluid velocity depends on the permeability and the pressure gradient , so the tissue is seen as a continuum with a certain resistance to flow rather than an architected mesh.

$$\bar{\tau} = \frac{\mu Q}{A\sqrt{K_p}}$$

$$K_p = \frac{\mu Q}{A\Delta p} h$$

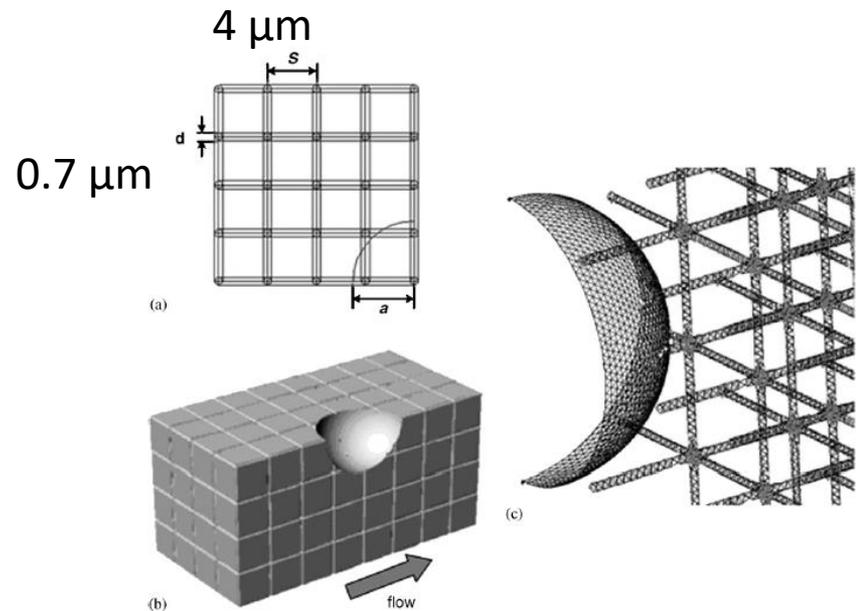
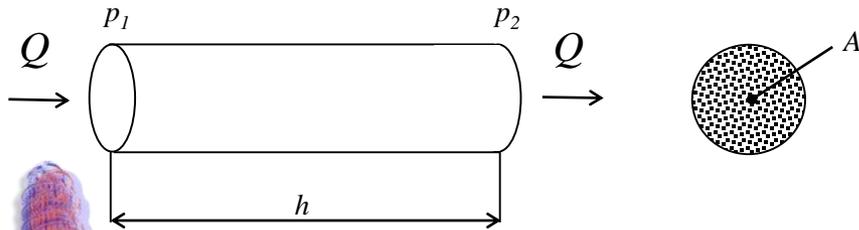


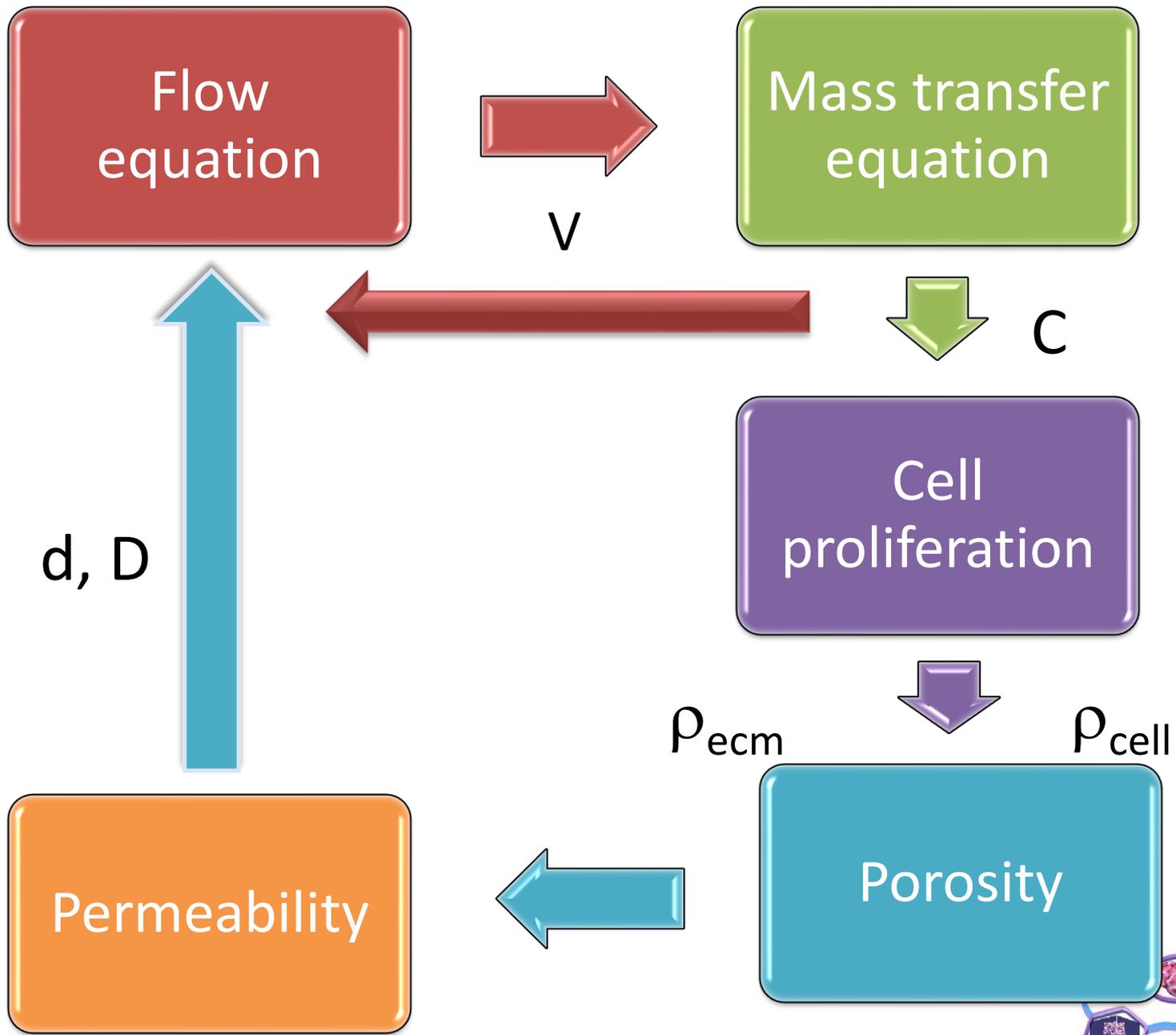
Fig. 1. CFD model setup. (a) Definition of the geometrical parameters of the model. (b) 3D rendering of the flow domain. (c) Detail of the mesh on the cell surface and nearby fibers.



The Brinkman correction to Darcys' equation takes into account the no slip condition at the walls of pores. (Darcy computes the pressure and the Brinkman correction computes the resulting velocity field as a function of fluid viscosity and permeability.)

$$q = \frac{-k}{\mu} \nabla p$$

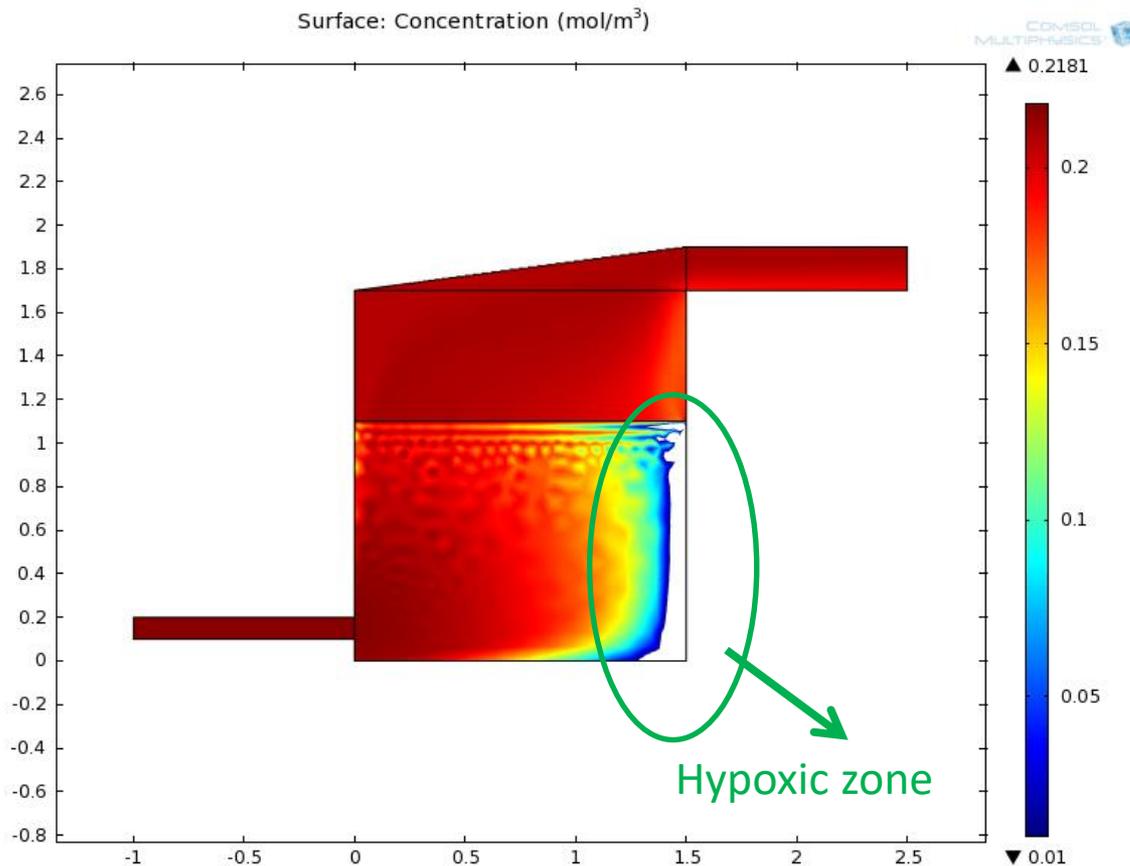
$$\mu \nabla^2 u + u = -k \nabla p$$



Oxygen consumption

Simulation

Adding reaction, and diffusion, convection multiphysics.
Sponge seeded with hepatocytes.



Reaction type Zero =>
constant consumption

Cell Density => $2.5 \cdot 10^{-6}$
cells/cm³

Hypoxic limit for hepatocyte
=> 0.01 mol/m³

OCR	Km	C crit	Co	D in water	D in sphere
1.10^{-18} to 1.10^{-16} moles.cell ⁻¹ .s ⁻¹	1.10^{-3} mM (0.7 mmHg)	1.10^{-4} mM (0.07 mmHg)	0.2 mM	3.10^{-9} m ² .s ⁻¹	2.10^{-9} m ² .s ⁻¹
Medium height	δ (Heaviside)	Cell density in vivo	Vmax	Flow rates	
1 mm	flc1hs(c-0.02,0.0001)	$8.8.10^{14}$ cells.m ⁻³	0.034 mM.s ⁻¹	10 to 500 μ L/min	

Cell density in body: 1.10^9 cells/mL

Cell density in vitro: 1.10^6 cells/mL